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Cell functions impaired by frataxin deficiency are restored by drug-mediated iron relocation

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Abstract

Various human disorders are associated with misdistribution of iron within or across cells. Friedreich ataxia (FRDA), a deficiency in the mitochondrial iron-chaperone frataxin, results in defective use of iron and its misdistribution between mitochondria and cytosol. We assessed the possibility of functionally correcting the cellular properties affected by frataxin deficiency with a siderophore capable of relocating iron and facilitating its metabolic use. Adding the chelator deferiprone at clinical concentrations to inducibly frataxin-deficient HEK-293 cells resulted in chelation of mitochondrial labile iron involved in oxidative stress and in reactivation of iron-depleted aconitase. These led to (1) restoration of impaired mitochondrial membrane and redox potentials, (2) increased adenosine triphosphate production and oxygen consumption, and (3) attenuation of mitochondrial DNA damage and reversal of hypersensitivity to staurosporine-induced apoptosis. Permeant chelators of higher affinity than deferiprone were not as efficient in restoring affected functions. Thus, although iron chelation might protect cells from iron toxicity, rendering the chelated iron bioavailable might underlie the capacity of deferiprone to restore cell functions affected by frataxin deficiency, as also observed in FRDA patients. The siderophore-like properties of deferiprone provide a rational basis for treating diseases of iron misdistribution, such as FRDA, anemia of chronic disease, and X-linked sideroblastic anemia with ataxia.

Topics: [aconitate hydratase](#), [iron](#), [mitochondria](#), [deferiprone](#), [chelating agents](#), [cytosol](#), [iron chelating agents](#), [dna](#), [mitochondrial](#), [oxidation-reduction](#), [adenosine triphosphate](#)

References

References

1

Weiss G, Goodnough LT. Anemia of chronic disease., *N Engl J Med*, 2005, vol. 352 (pg. 1011-1023)

[Google Scholar](#) [Crossref](#) [PubMed](#)

2

Ganz T. Molecular pathogenesis of anemia of chronic disease., *Pediatr Blood Cancer*, 2006, vol. 46 (pg. 554-557)

[Google Scholar](#) [Crossref](#) [PubMed](#)

3

Delatycki MB, Williamson R, Forrest SM. Friedreich ataxia: an overview., *J Med Genet*, 2000, vol. 37 (pg. 1-8)

4

Patel PI, Isaya G. Friedreich ataxia: from GAA triplet-repeat expansion to frataxin deficiency, *Am J Hum Genet*, 2001, vol. 69 (pg. 15-24)

[Google Scholar](#) [Crossref](#) [PubMed](#)

5

Rotig A, de Lonlay P, Chretien D, et al. Aconitase and mitochondrial iron-sulphur protein deficiency in Friedreich ataxia., *Nat Genet*, 1997, vol. 17 (pg. 215-217)

[Google Scholar](#) [Crossref](#) [PubMed](#)

6

Honda K, Casadesus G, Petersen RB, Perry G, Smith MA. Oxidative stress and redox-active iron in Alzheimer's disease., *Ann N Y Acad Sci*; 2004, vol. 1012 (pg. 179-182)

[Google Scholar](#) [Crossref](#) [PubMed](#)

7

Youdim MBH, Stephenson G, Shachar DB. Ironing iron out in Parkinson's disease and other neurodegenerative diseases with iron chelators: a lesson from 6-hydroxydopamine and iron chelators, desferal and VK-28., *Ann N Y Acad Sci*; 2004, vol. 1012 (pg. 306-325)

[Google Scholar](#) [Crossref](#) [PubMed](#)

8

Pandolfo M. Friedreich ataxia., *Semin Pediatr Neurol*, 2003, vol. 10 (pg. 163-172)

[Google Scholar](#) [Crossref](#) [PubMed](#)

9

Bulteau A-L, Dancis A, Gareil M, Montagne J-J, Camadro J-M, Lesuisse E. Oxidative stress and protease dysfunction in the yeast model of Friedreich ataxia., *Free Radic Biol Med*, 2007, vol. 42 (pg. 1561-1570)

[Google Scholar](#) [Crossref](#) [PubMed](#)

10

Waldvogel D, van Gelderen P, Hallett M. Increased iron in the dentate nucleus of patients with Friedrich's ataxia., *Ann Neurol*, 1999, vol. 46 (pg. 123-125)

[Google Scholar](#) [Crossref](#) [PubMed](#)

11

Gakh O, Park S, Liu G, et al. Mitochondrial iron detoxification is a primary function of frataxin that limits oxidative damage and preserves cell longevity., *Hum Mol Genet*, 2006, vol. 15 (pg. 467-479)

[Google Scholar](#) [Crossref](#) [PubMed](#)

12

Cossee M, Puccio H, Gansmuller A, et al. Inactivation of the Friedreich ataxia mouse gene leads to early embryonic lethality without iron accumulation., *Hum Mol Genet*, 2000, vol. 9 (pg. 1219-1226)

[Google Scholar](#) [Crossref](#) [PubMed](#)

Abstract



Introduction

13

Methods

Puccio H, Simon D, Cossee M, et al. Mouse models for Friedreich ataxia exhibit cardiomyopathy, sensory nerve defect and Fe-S enzyme deficiency followed by intramitochondrial iron deposits., *Nat Genet*, 2001, vol. 27 (pg. 181-186)

Results

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[Google Scholar](#) [Crossref](#) [PubMed](#)

Acknowledgments

Stehling O, Elense H-P, Bruckel B, Muhlenhoff U, Lill R. Iron-sulfur protein maturation in human cells: evidence for a function of frataxin., *Hum Mol Genet*, 2004, vol. 13 (pg. 3007-3015)

[Google Scholar](#) [Crossref](#) [PubMed](#)

15

Sturm B, Bistrich U, Schranzhofer M, et al. Friedreich's ataxia, no changes in mitochondrial labile iron in human lymphoblasts and fibroblasts: a decrease in antioxidative capacity?, *J Biol Chem*, 2005, vol. 280 (pg. 6701-6708)

[Google Scholar](#) [Crossref](#) [PubMed](#)

16

Kalinowski DS, Richardson DR. The evolution of iron chelators for the treatment of iron overload disease and cancer., *Pharmacol Rev*, 2005, vol. 57 (pg. 547-583)

[Google Scholar](#) [Crossref](#) [PubMed](#)

17

Sohn Y-S, Breuer W, Munnich A, Cabantchik ZI. Redistribution of accumulated cell iron: a modality of chelation with therapeutic implications., *Blood*, 2008, vol. 111 (pg. 1690-1699)

[Google Scholar](#) [Crossref](#) [PubMed](#)

18

Fredenburg AM, Sethi RK, Allen DD, A. Yokel R. The pharmacokinetics and blood-brain barrier permeation of the chelators 1,2 dimethyl-, 1,2 diethyl-, and 1-[ethan-1' ol]-2-methyl-3-hydroxypyridin-4-one in the rat., *Toxicology*, 1996, vol. 108 pg. 191

[Google Scholar](#) [Crossref](#) [PubMed](#)

19

Boddaert N, Le Quan Sang KH, Rotig A, et al. Selective iron chelation in Friedreich ataxia: biologic and clinical implications., *Blood*, 2007, vol. 110 (pg. 401-408)

[Google Scholar](#) [Crossref](#) [PubMed](#)

20

Lu C, Cortopassi G. Frataxin knockdown causes loss of cytoplasmic iron-sulfur cluster functions, redox alterations and induction of heme transcripts., *Arch Biochem Biophys*, 2007, vol. 457 (pg. 111-122)

[Google Scholar](#) [Crossref](#) [PubMed](#)

21

Kingston RE, Chen CA, Rose JK. Unit 9.1: Calcium Phosphate Transfection., *Current Protocols in Molecular Biology*, 2003Hoboken, NJJohn Wiley & Sons

[Google Scholar](#)

22

Glickstein H, Ben El R, Shvartsman M, Cabantchik ZI. Intracellular labile iron pools as direct targets of iron chelators: a fluorescence study of chelator action in living cells., *Blood*, 2005, vol. 106 (pg. 3242-3250)

[Google Scholar](#) [Crossref](#) [PubMed](#)

23

Schwarzer C, Illek B, Suh JH, Remington SJ, Fischer H, Machen TE. Organelle redox of CF and CFTR-corrected airway epithelia., *Free Radic Biol Med*, 2007, vol. 43 pg. 300

[Google Scholar](#) [Crossref](#) [PubMed](#)

24

Epsztejn S, Kakhlon O, Glickstein H, Breuer W, Cabantchik ZI. Fluorescence analysis of the labile iron pool of mammalian cells., *Anal Biochem*, 1997, vol. 248 (pg. 31-42)

[Google Scholar](#) [Crossref](#) [PubMed](#)

25

Drapier JC, Hibbs JB. Aconitases: a class of metalloproteins highly sensitive to nitric oxide synthesis, *Methods Enzymol*, 1996, vol. 269 (pg. 26-36)

[Google Scholar](#) [Crossref](#) [PubMed](#)

26

Tong W-H, Rouault TA. Functions of mitochondrial ISCU and cytosolic ISCU in mammalian iron-sulfur cluster biogenesis and iron homeostasis., *Cell Metab*, 2006, vol. 3 (pg. 199-210)

[Google Scholar](#) [Crossref](#) [PubMed](#)

27

Glickstein H, El RB, Link G, et al. Action of chelators in iron-loaded cardiac cells: accessibility to intracellular labile iron and functional consequences., *Blood*, 2006, vol. 108 (pg. 3195-3203)

[Google Scholar](#) [Crossref](#) [PubMed](#)

28

Li K, Besse EK, Ha D, Kovtunovych G, Rouault TA. Iron-dependent regulation of frataxin expression: implications for treatment of Friedreich ataxia., *Hum Mol Genet*, 2008, vol. 17 (pg. 2265-2273)

[Google Scholar](#) [Crossref](#) [PubMed](#)

29

Babcock M, de Silva D, Oaks R, et al. Regulation of mitochondrial iron accumulation by Yfh1p, a putative homolog of frataxin., *Science*, 1997, vol. 276 (pg. 1709-1712)

[Google Scholar](#) [Crossref](#) [PubMed](#)

30

Kispal G, Csere P, Prohl C, Lill R. The mitochondrial proteins Atm1p and Nfs1p are essential for biogenesis of cytosolic Fe/S proteins, *EMBO J*, 1999, vol. 18 (pg. 3981-3989)

[Google Scholar](#) [Crossref](#) [PubMed](#)

31

Foury F, Talibi D. Mitochondrial control of iron homeostasis: a genome wide analysis of gene expression in a yeast frataxin-deficient strain, *J Biol Chem*, 2001, vol. 276 (pg. 7762-7768)

[Google Scholar](#) [Crossref](#) [PubMed](#)

32

Hausmann A, Samans B, Lill R, Muhlenhoff U. Cellular and mitochondrial remodelling upon defects in iron-sulfur protein biogenesis., *J Biol Chem*, 2008, vol. 283 (pg. 8318-8330)

[Google Scholar](#) [Crossref](#) [PubMed](#)

33

Garrido C, Galluzzi L, Brunet M, Puig PE, Didelot C, Kroemer G. Mechanisms of cytochrome c release from mitochondria., *Cell Death Differ*, 2006, vol. 13 (pg. 1423-1433)

[Google Scholar](#) [Crossref](#) [PubMed](#)

34

Detmer SA, Chan DC. Functions and dysfunctions of mitochondrial dynamics., *Nat Rev Mol Cell Biol*, 2007, vol. 8 (pg. 870-879)

[Google Scholar](#) [Crossref](#) [PubMed](#)

35

Napoli E, Taroni F, Cortopassi GA. Frataxin, iron-sulfur clusters, heme, ROS, and aging., *Antioxidants Redox Signal*, 2006, vol. 8 (pg. 506-516)

[Google Scholar](#) [Crossref](#)

36

Wu SP, Cowan JA. Iron-sulfur cluster stability: kinetics and mechanism of ligand-promoted cluster degradation, *Chem Commun (Camb)*, 2007, vol. 1 (pg. 82-84)

[Google Scholar](#) [Crossref](#)

37

Foury F. Low iron concentration and aconitase deficiency in a yeast frataxin homologue deficient strain., *FEBS Lett*, 1999, vol. 456 pg. 281

[Google Scholar](#) [Crossref](#) [PubMed](#)

38

Rouan MC, Marfil F, Mangoni P, Sechaud R, Humbert H, Maurer G. Determination of a new oral iron chelator, ICL670, and its iron complex in plasma by high-performance liquid chromatography and ultraviolet detection., *J Chromatogr B Biomed Sci Appl*, 2001, vol. 755 (pg. 203-213)

[Google Scholar](#) [Crossref](#) [PubMed](#)

39

Rouault TA, Cooperman S. Brain iron metabolism., *Semin Pediatric Neurol*, 2006, vol. 13 (pg. 142-148)

[Google Scholar](#) [Crossref](#)

40

Theurl I, Mattle V, Seifert M, Mariani M, Marth C, Weiss G. Dysregulated monocyte iron homeostasis and erythropoietin formation in patients with anemia of chronic disease., *Blood*, 2006, vol. 107 (pg. 4142-4148)

[Google Scholar](#) [Crossref](#) [PubMed](#)

41

Napier I, Ponka P, Richardson DR. Iron trafficking in the mitochondrion: novel pathways revealed by disease., *Blood*, 2005, vol. 105 (pg. 1867-1874)

[Google Scholar](#) [Crossref](#) [PubMed](#)

42

Zanella I, Derosas M, Corrado M, et al. The effects of frataxin silencing in HeLa cells are rescued by the expression of human mitochondrial ferritin., *Biochim Biophys Acta*, 2008, vol. 1782 (pg. 90-98)

[Google Scholar](#) [Crossref](#) [PubMed](#)

43

Vreugdenhil G, Swaak AJ, de Jeu-Jaspers C, van Eijk HG. Correlation of iron exchange between the oral iron chelator 1,2-dimethyl-3-hydroxypyrid-4-one(L1) and transferrin and possible antianaemic effects of L1 in rheumatoid arthritis., *Ann Rheum Dis*, 1990, vol. 49 (pg. 956-957)

[Google Scholar](#) [Crossref](#) [PubMed](#)

44

Oktay Y, Dioum E, Matsuzaki S, et al. Hypoxia-inducible factor 2 α regulates expression of the mitochondrial aconitase chaperone protein frataxin., *J Biol Chem*, 2007, vol. 282 (pg. 11750-11756)

[Google Scholar](#) [Crossref](#) [PubMed](#)

45

Sturm B, Stupphann D, Kaun C, et al. Recombinant human erythropoietin: effects on frataxin expression in vitro., *Eur J Clin Invest*, 2005, vol. 35 (pg. 711-717)

[Google Scholar](#) [Crossref](#) [PubMed](#)

46

Wang GL, Semenza GL. Desferrioxamine induces erythropoietin gene expression and hypoxia-inducible factor 1 DNA-binding activity: implications for models of hypoxia signal transduction., *Blood*, 1993, vol. 82 (pg. 3610-3615)

[Google Scholar](#) [Crossref](#) [PubMed](#)

47

Cohen AR, Galanello R, Piga A, De Sanctis V, Tricta F. Safety and effectiveness of long-term therapy with the oral iron chelator deferiprone., *Blood*, 2003, vol. 102 (pg. 1583-1587)

[Google Scholar](#) [Crossref](#) [PubMed](#)

48

Forni GL, Balocco M, Cremonesi L, Abbruzzese G, Parodi RC, Marchese R. Regression of symptoms after selective iron chelation therapy in a case of neurodegeneration with brain iron accumulation., *Mov Disord*, 2008, vol. 23 (pg. 904-907)

[Google Scholar](#) [Crossref](#) [PubMed](#)

49

Liu G, Hider R, Templeton D. Iron chelator chemistry., *Molecular and Cellular Iron Transport*, 2002 New York, NY Marcel Dekker pg. 321

[Google Scholar](#)

50

El-Jammal A, Templeton DM. Iron-hydroxypyridone redox chemistry: kinetic and thermodynamic limitations to Fenton activity., *Inorgan Chim Acta*, 1996, vol. 245 pg. 199

[Google Scholar](#) [Crossref](#)

51

Konijn AM, Glickstein H, Vaisman B, Meyron-Holtz EG, Slotki IN, Cabantchik ZI. The cellular labile iron pool and intracellular ferritin in K562 cells., *Blood*, 1999, vol. 94 (pg. 2128-2134)

[Google Scholar](#) [Crossref](#) [PubMed](#)

52

Goncalves S, Paupe V, Dassa E, Rustin P. Deferiprone targets aconitase: implication for Friedreich's ataxia treatment., *BMC Neuro*, 2008, vol. 8 pg. 20

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Drugs Used to Treat Iron Deficiency Anemia. The following list of medications are in some way related to, or used in the treatment of this condition. Select drug class. All drug classes - iron products (51) - vitamin and mineral combinations (7) - phosphate binders (2). Rx. OTC. Brand names: Feosol Original, Slow Fe, FeroSul, Fer-In-Sol, Slow Release Iron, Ferrousal, Fer-Iron, Fe Caps, Fer-Gen-Sol, Feratab, Ferra T.D. Caps, Ferro-Bob ...show all. Drug class: iron products. For consumers: dosage, interactions, For professionals: A-Z Drug Facts, Prescribing Information. The function of frataxin is not clear but it is involved in assembly of iron-sulfur clusters. It has been proposed to act as either an iron chaperone or an iron storage protein. Reduced expression of frataxin is the cause of Friedreich's ataxia. Contents. Frataxin mRNA is predominantly expressed in tissues with a high metabolic rate (including liver, kidney, brown fat and heart). Mouse and yeast frataxin homologues contain a potential N-terminal mitochondrial targeting sequence, and human frataxin has been observed to co-localise with a mitochondrial protein. Furthermore, disruption of the yeast gene has been shown to result in mitochondrial dysfunction.